

T cell cytotoxicity

[Immunotherapy](#) has transformed [cancer treatments](#); however, a large fraction of [patients](#) encounter [resistance](#). Such resistance is mediated by complex factors, often involving interactions between multiple [genes](#). Thus, it is crucially important to identify genetic interactions between genes that are significantly mutated in cancer patients and those involved in [immune responses](#), ideally the ones that currently have chemical compounds for direct targeting. To systematically interrogate such genetic interactions that mediate cancer cells' response to [T cell](#) killing, Park et al. designed an asymmetric dual perturbation [library](#) targeting the matched combinations between significantly mutated [tumor suppressors](#) and immune resistance genes. They performed a combinatorial double [knockout](#) screen on 1159 gene pairs and identified those where joint loss-of-function renders altered cellular response to [T cell cytotoxicity](#). They also performed comparative [transcriptomics](#)-based analyses on tumor and normal samples from [TCGA](#) and [Genotype-Tissue Expression](#) cohorts, [mutational](#) profiling analyses, and [survival analysis](#) to further characterize the significance of identified hits in clinical patients. Interactions between significantly mutated [tumor suppressors](#) and potentially druggable [immune resistance](#) genes may offer insights on potential new concepts of how to target clinically relevant cancer mutations with currently available agents ¹⁾

¹⁾

Park JJ, Codina A, Ye L, Lam S, Guo J, Clark P, Zhou X, Peng L, Chen S. Double knockout CRISPR screen for cancer resistance to T cell [cytotoxicity](#). J Hematol Oncol. 2022 Dec 1;15(1):172. doi: 10.1186/s13045-022-01389-y. PMID: 36456981.

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