The ability to rapidly assess and monitor patient immune responses is critical for clinical diagnostics; vaccine design; and fundamental investigations into the presence or generation of protective immunity against infectious diseases. Findings on the limits of antibody-based protection provided by B-cells have highlighted the importance of engaging pathogen-specific T-cells for long-lasting and broad protection against viruses and their emergent variants such as in SARS-CoV-2. However; lowcost and point-of-care tools for detecting engagement of T-cell immunity in patients are conspicuously lacking in ongoing efforts to assess and control population-wide disease risk. Currently available tools for human T-cell analysis are time and resource-intensive. Utilizing multichannel silicon nanowire field-effect transistors (Si-NW-FET) compatible with complementary metal-oxide-semiconductor (CMOS); we developed a device designed for rapid and label-free detection of human T-cell immune responses. We demonstrate the generalizability of this approach by measuring T-cell responses against melanoma antigen MART1; common and seasonal viruses CMV; EBV; flu; as well as emergent pandemic coronavirus; SARS-CoV-2. Further; this device provides a modular and translational platform for optimizing vaccine formulations and combinations; offering guick and guantitative readouts for acquisition and persistence of T-cell immunity against variant-driven pathogens such as Flu and pandemic SARS-CoV-2¹⁾

In a issue of Neuron, Stoeber et al. (2018) report a biosensor resolving the spatiotemporal organization of opioid receptor activation in living neurons. They delineate novel signaling mechanisms in endosomes and Golgi differentially engaged by opioid peptides and drugs ²⁾.

1)

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