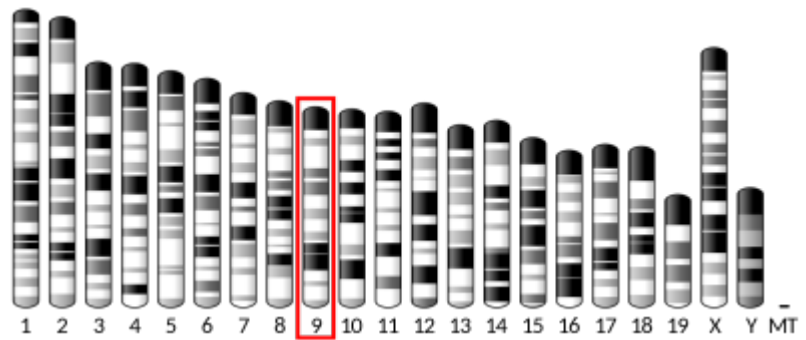


# BBS9 gene in nonsyndromic craniosynostosis

Mutations in several genes account for a small number of Nonsyndromic craniosynostosis (NCS) patients; thus, the molecular etiopathogenesis of NCS remains largely unclear.



In 2012 Justice et al., conducted, the first genome-wide association study for nonsyndromic sagittal craniosynostosis (sNSC) using 130 non-Hispanic case-parent trios of European ancestry (NHW). They found robust associations in a 120-kb region downstream of *BMP2* flanked by rs1884302 ( $P = 1.13 \times 10^{-14}$ , odds ratio (OR) = 4.58) and rs6140226 ( $P = 3.40 \times 10^{-11}$ , OR = 0.24) and within a 167-kb region of *BBS9* between rs10262453 ( $P = 1.61 \times 10^{-10}$ , OR = 0.19) and rs17724206 ( $P = 1.50 \times 10^{-8}$ , OR = 0.22). They replicated the associations to both loci (rs1884302,  $P = 4.39 \times 10^{-31}$ ) and rs10262453,  $P = 3.50 \times 10^{-14}$ ) in an independent NHW population of 172 unrelated probands with sNSC and 548 controls. Both *BMP2* and *BBS9* are genes with roles in skeletal development that warranted functional studies to further understand the etiology of sNSC <sup>1)</sup>.

Sewda et al., identified a novel *BBS9* variant that further shows the potential involvement of *BBS9* in the pathogenesis of craniosynostosis (CS) <sup>2)</sup>

A study of Barba et al., aimed at characterizing the molecular signaling implicated in the aberrant ossification of cranial sutures in NCS patients. Comparative gene expression profiling of NCS patient sutures identified a fused suture-specific signature, including 17 genes involved in primary cilium signaling and assembly. Cells from fused sutures displayed a reduced potential to form primary cilia compared to cells from control patent sutures of the same patient.

They identified specific upregulated splice variants of the Bardet Biedl syndrome-associated gene 9 (*BBS9*), which encodes a structural component of the ciliary BBSome complex. *BBS9* expression increased during in vitro osteogenic differentiation of suture-derived mesenchymal cells of NCS patients. Also, *Bbs9* expression increased during in vivo ossification of rat sutures. *BBS9* functional knockdown affected the expression of primary cilia on patient suture cells and their osteogenic potential. Computational modeling of the upregulated protein isoforms (observed in patients) predicted that their binding affinity within the BBSome may be affected, providing a possible explanation for the aberrant suture ossification in NCS <sup>3)</sup>.

Previous genome-wide association study of sagittal nonsyndromic craniosynostosis identified associations with variants downstream from *BMP2* and intronic in *BBS9*. Because no coding variants in *BMP2* were identified, Justice et al., hypothesized that conserved non-coding regulatory elements may alter *BMP2* expression. In order to identify and characterize noncoding regulatory elements near

BMP2, two conserved noncoding regions near the associated region on chromosome 20 were tested for regulatory activity with a Renilla **luciferase** assay. For a 711 base pair noncoding fragment encompassing the most strongly associated variant, rs1884302, the **luciferase** assay showed that the risk allele (C) of rs1884302 drives higher expression of the reporter than the common allele (T). When this same DNA fragment was tested in zebrafish transgenesis studies, a strikingly different expression pattern of the green fluorescent reporter was observed depending on whether the transgenic fish had the risk (C) or the common (T) allele at rs1884302. The in vitro results suggest that altered BMP2 regulatory function at rs1884302 may contribute to the etiology of sagittal nonsyndromic craniosynostosis. The in vivo results indicate that differences in regulatory activity depend on the presence of a C or T allele at rs1884302 <sup>4)</sup>.

## References

1)

Justice CM, Yagnik G, Kim Y, Peter I, Jabs EW, Erazo M, Ye X, Ainehsazan E, Shi L, Cunningham ML, Kimonis V, Roscioli T, Wall SA, Wilkie AO, Stoler J, Richtsmeier JT, Heuzé Y, Sanchez-Lara PA, Buckley MF, Druschel CM, Mills JL, Caggana M, Romitti PA, Kay DM, Senders C, Taub PJ, Klein OD, Boggan J, Zwienerberg-Lee M, Naydenov C, Kim J, Wilson AF, Boyadjiev SA. A genome-wide association study identifies susceptibility loci for nonsyndromic sagittal craniosynostosis near BMP2 and within BBS9. *Nat Genet.* 2012 Dec;44(12):1360-4. doi: 10.1038/ng.2463. Epub 2012 Nov 18. PubMed PMID: 23160099; PubMed Central PMCID: PMC3736322.

2)

Sewda A, White SR, Erazo M, Hao K, García-Fructuoso G, Fernández-Rodríguez I, Heuzé Y, Richtsmeier JT, Romitti PA, Reva B, Jabs EW, Peter I. Nonsyndromic craniosynostosis: novel coding variants. *Pediatr Res.* 2019 Mar;85(4):463-468. doi: 10.1038/s41390-019-0274-2. Epub 2019 Jan 14. PubMed PMID: 30651579; PubMed Central PMCID: PMC6398438.

3)

Barba M, Di Pietro L, Massimi L, Geloso MC, Frassanito P, Caldarelli M, Michetti F, Della Longa S, Romitti PA, Di Rocco C, Arcovito A, Parolini O, Tamburrini G, Bernardini C, Boyadjiev SA, Lattanzi W. BBS9 gene in nonsyndromic craniosynostosis: Role of the primary cilium in the aberrant ossification of the suture osteogenic niche. *Bone.* 2018 Jul;112:58-70. doi: 10.1016/j.bone.2018.04.013. Epub 2018 Apr 17. Erratum in: *Bone.* 2019 Apr;121:293. PubMed PMID: 29674126; PubMed Central PMCID: PMC5970090.

4)

Justice CM, Kim J, Kim SD, Kim K, Yagnik G, Cuellar A, Carrington B, Lu CL, Sood R, Boyadjiev SA, Wilson AF. A variant associated with sagittal nonsyndromic craniosynostosis alters the regulatory function of a non-coding element. *Am J Med Genet A.* 2017 Nov;173(11):2893-2897. doi: 10.1002/ajmg.a.38392. Epub 2017 Oct 6. PubMed PMID: 28985029; PubMed Central PMCID: PMC5659764.

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